



PROFICIENCY TESTING REPORT

ISHTM-AIIMS EXTERNAL QUALITY ASSURANCE PROGRAMME NABL accredited program as per ISO/IEC 17043:2010 standard Organized By Department of Hematology, AIIMS, New Delhi-110029

Duration of stability testing - minimum upto 8 days at ambient temp. after dispatch of specimens

EQAP CODE No. : 1702

Distribution No.: 151-D

Month/Year: August/2020

Instrument ID: MEK 6510K

Name & Contact No. of PT Co-ordinator: Dr. Seema Tyagi (Prof.), Hematology, AIIMS, Delhi,

Tel: 9013085730, E-Mail: accuracy2000@gmail.com Date of issue & status of the report: 27-11-2020[Final].

CBC and Retic Assessment

			Among Lab (Accuracy Testing)					Within Lab (Precision Testing)				
Test Parameters	S.No.	Your Result 1			Consensus result	Uncertainty of Assigned Values		Yours Results	Consensus Result Diff. of 2 values (Assigned Value)		7	
WBC x10³/μl	1	5.3	5.1	10.4	9.8	0.0460	0.43	0.2	0.1	0.0060	0.96	
RBC x10 ⁶ /µl	1	3.17	3.07	6.24	6.4	0.0060	-0.90	0.1	0.03	0.0010	1.89	
Hb g/dl	1	11.4	11.1	22.5	23.4	0.0210	-1.52	0.3	0.1	0.0070	2.57	
HCT%	1	34.9	34.1	69	72.05	0.1860	-0.50	0.8	0.3	0.0200	1.35	
MCV-fl	1	111	110	221	223.5	0.4950	-0.14	1	0.3	0.0230	1.57	
MCH-Pg	1	36.2	36	72.2	73.2	0.0760	-0.48	0.2	0.3	0.0200	-0.34	
MCHC-g/dl	1	32.7	32.6	65.3	65.3	0.1640	0.00	0.1	0.3	0.0200	-0.67	
Plt. x10³/μl	1	241	228	469	512	1.38	-1.12	13	6	0.32	1.22	
Retic %	2	2.7	2.6	5.3	7.2	0.13	-0.51	0.1	0.3	0.02	-0.72	

P.S. Assesment

		YOUR REPORT	CONSENSUS REPORT				
DLC%	3	Nrbcs=0-1/hpf , Poly=22% L=04%, E=00%, Mono/Promono=0% , B1=11% P.M.=17%, Mye=28%, Meta=18%, Other=	Poly: 25-50, Lymph; 2-7, nRBC/Mono/Eo/Blast/Pro: 0-5, Myelo: 20-35, Meta: 15-25, Baso: 0-3				
RBC Morphology	3	Mild hypochromasia with Mainly Macrocytic and few Microcytic Rbc with mod. Anisocytosis and mild poikilocytosis with few tear drop shaped rbc and few fragmented rbc seen.	Predominantly: Normocytic Normochromic. Moderate: Anisocytosis. Mild: Microcytic				
Diagnosis	3	Chronic Myeloid Leukaemia	Chronic Myeloid Leukemia (Chronic Phase) : CML-CP				

COMBINED DATA VALUES OF TOTAL PARTICIPANTS

		77		T					tab 7
Test parameters	S.No.	Total participants covered in	Total No. responded	% of Labs with Z Score 0-2		% of Labs with Z Score 2-3		Score >3	
	0.110.	the current dist.		Among labs	Within lab	Among labs	Within lab	Among labs	Within lab
WBC $x10^3/\mu l$	1	450	363	88.43	86.23	4.68	5.79	6.61	7.71
RBC x10 ⁶ /μl	1	450	363	88.43	88.98	4.96	6.06	6.34	4.68
Hb g/dl	1	450	363	85.12	90.91	6.34	4.41	8.54	4.68
HCT%	1	450	363	94.77	92.29	3.03	2.75	1.93	4.68
MCV-fl	1	450	363	97.8	92.29	0.55	3.03	1.1	4.13
MCH-Pg	1	450	363	88.98	90.91	5.51	5.23	4.96	3.03
MCHC-g/dl	1	450	363	96.97	87.05	1.38	6.06	1.38	6.06
Plt. x10 ³ /µl	1	450	363	90.08	90.63	5.23	5.51	4.41	3.58
ReticCount%	2	450	322	93.17	83.85	4.35	1.55	2.8	16.15
PS Assessment	3	450	352	Acceptable:75.4%, Warning Signal:24.6%, Unacceptable:0%					

'Comments:

- 1). Among Lab (EQA): Results acceptable.
- 2). Within Lab (IQA): Precision acceptable.

Note-1: EQA (External Quality Assurance): Your Performance among various of participating labs in PT, to determine the accuracy of your results.

IQA (Internal Quality Assurance): Your Performance of comparison of two consecutive measurement values within your lab to test the precision of your autoanalyzer.

Note-2: Z score among & within lab were calculated, as per to ISO/IEC 13528:2015 standard. Z score among lab (EQA)= (Your Result Sum of two values - Consensus Result sum of two values)/(Normalised IQR)

Z score within lab (IQA)= (Your Result Difference of two values - Consensus Result difference of two values)/(Normalised IQR)

IQR = Quartile 3 - Quartile 1 of participant data, Normalised $IQR = 0.7413 \times IQR$

Note-3: Z score 0 to ± 2 : Acceptable, Z score ± 2 to ± 3 : Warning Signal, Z score > ± 3 : Unacceptable [As per ISO/IEC 13528:2015 standard]

Note-4: Z score value between "0 to ± 2 " are texted in green colour. Z score value between " ± 2 to ± 3 " are texted in orange colour. Z score value > ± 3 are texted in red colour.

Note-5: Homogeneity and stability testing of PT sample were done as per ISO 13528:2015 standard. To pass homogeneity test, between sample SD (Ss) should be smaller than the check value (0.3*SDPA). To pass the stability test, average difference in measurement values of first and last day sample $(\bar{x}-\bar{y})$ should be smaller than the check value (0.3*SDPA).

Note-6: ISHTM-AIIMS-EQAP does not subcontract any task of its scheme

Note-7: Participants are free to use methods/analyzer of their own choice.

Note-8: Proficiency testing (PT) samples are sent quarterly to each participant.

Note-9: All the necessary details regarding design and implementation of PT, are provided in the instruction sheet as well as on programme's website www.ishtmaiimseqap.com.

Report authorized by,

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Dr. Seema Tyagi (Prof.)

PT Co-ordinator: ISHTM-AIIMS-EQAP

Department of Hematology, AIIMS, New Delhi

-----End Of Report-----

Re: M(EL)T Application - M(EL)T-00064

ALAKNANDA DIAGNOSTIC LAB <alaknandadiagnosticlab@gmail.com>

Mon 2/22/2021 12:24 PM

To: NABL M(EL)T <nablmelt@nabl.qcin.org>

1 attachments (10 MB)

CamScanner 11-18-2020 14.45.30.pdf;

Dear Sir,

For Haematology equas August report is the last sample and report I received as due to Covid few samples were missed in between for the AIIMS equas which can be checked from AIIMS. For 2021 I have received an equas sample in February for which report will come in March. I am attaching the raw data for the calibration of the Nihon Kohden machine.

Thanks and Regards. Dr Rohini Kalhan Alaknanda Diagnostic Lab.

On Mon, 22 Feb 2021, 11:58 NABL M(EL)T, < nablmelt@nabl.qcin.org> wrote: Dear Madam,

This mail is to inform you that August 2020 EQAS result has been attached. Please send the latest EQAS result of Haematology.

Also, please send the raw data that has been observed during calibration for the Haematology analyser i.e. Nihon Kohden 6510K.

Sincerely,